

Short Communication

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Development of a murine model of metabolic syndrome for evaluation with hyperbaric oxygen therapy

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Abstract

The metabolic syndrome is a worldwide distributed clinical entity that significantly affects the functional state of the individual. The development of murine models for experimentation in this area has generally been associated with the use of drugs that generate irreversible acute damage to various organs. The objective of this study is to develop a murine model based on the sucrose administration for 20 weeks. Through this process, rats with weight gain, fat accumulation and biochemical parameters characteristic of metabolic syndrome were obtained for a subsequent evaluation with hyperbaric oxygen therapy.

Keywords

Wistar,
metabolic syndrome,
hyperbaric oxygen therapy,
sucrose

Non-standard abbreviations:

MetS: Metabolic syndrome
HBOT: Hyperbaric
Oxygen therapy

Introduction

The Metabolic Syndrome (MetS) is a clinical entity with an important impact worldwide that is characterized by overweight or obesity, insulin resistance and pro-inflammatory processes, whose consequences are not limited solely to the functional state of the individual who develops it, but also in the economic, social and family [1]. The presence of MS represents a notable increase in overweight and risk factors to develop type 2 diabetes mellitus, hypertriglyceridemia, increase in low density lipoprotein (LDL), decrease in high density lipoprotein (HDL), and systemic arterial hypertension. Conventional treatment alternatives for this clinical entity use different types of strategies such as drugs (statins, benzofibrates, oral hypoglycemic agents, among others), or modifications of dietary and hygienic habits, which have been repeatedly considered a therapeutic failure, since no epidemiological decrease of this clinical condition has been seen, but it is increasing [2].

Hyperbaric Oxygen Therapy (HBOT) is a therapy mainly indicated for the treatment of Decompressive Disease, Gaseous Aeroembolism and Carbon Monoxide Poisoning. There are other indications where benefits are appreciated for the patient, for example, as an adjuvant for scarring processes, refractory osteomyelitis, gas gangrene, just to mention a few. Of the conditions described above, the aid for scarring processes is undoubtedly the main reason why this treatment is used, being the lesions complicated by metabolic processes the main causes [3,4].

The development of murine models has historically shown great efficiency in basic research protocols, with advantages such as easy handling, low cost, easy reproduction and homogeneity of the study subjects, which represents a statistical advantage by reducing error factors. Commonly, the development of animal models with MetS involves the application of a drug that causes irreversible acute damage to the pancreas, liver or kidney [5].

With the aim of being able to evaluate the efficacy of treatment with HBOT in cases of MetS, in this work a murine model was fed with water with sucrose as an obesity inducer, developing fatty liver, increased blood pressure, tachycardia, alterations of the glycemia and significant weight gain [6], being clinically a better study model resembling the natural history of MetS.

Results and Discussion

Figure 1 shows the evolution of the body weight of rats that consumed sucrose during 20 weeks; rats in the Metabolic syndrome groups that ingested sucrose (MetS group) showed significant changes in the body weight with respect to the control group ($P < 0.05$). From week 1 to week 14 there was a steady increase in the body weight of the MetS group with respect to the control group. These results are consistent with those reported by other authors who have used sucrose or fructose as dietary inducers of metabolic disorders in rats, such as insulin resistance, obesity and MetS [7,8]. Experimental evidence shows that the consumption of sucrose increases the caloric intake and therefore the body weight; the mechanism implicated has been related with the consumption of sugars (sucrose or fructose) in liquid form, both short [9] and long term [10] that is associated with a lower satiety due to the inability to stimulate the production of leptin and others hormones tending to increase the caloric intake during subsequent meals. [11].

After 20 weeks of sucrose intake, rats in the MetS group showed a significant increase in body weight of 38% ($P < 0.01$), total fat (84%, $P < 0.05$), adiposity index (33%, $P < 0.05$) and triglycerides (95%, $P < 0.05$), while HDL decreased 48% ($P < 0.05$). Finally, no difference was observed in the levels of serum glucose and caloric intake. All the results above were compared with respect to the control group (Table 1). In relation to the above, the MetS group developed a large amount of abdominal and epididymal fat. Regarding this phenomenon of adipose tissue storage, some authors have reported that the accumulation of intramuscular and visceral fat is a primary factor in the pathogenesis of a variety of metabolic disorders, including insulin resistance, type 2 diabetes and hyperlipidemia [12,13], but mainly, abdominal obesity that develops MetS and non-alcoholic fatty liver [14,15]. In humans, adipose tissue produces and secretes a large number of pro-inflammatory adipocytokines that causes an imbalance in the secretion of interleukins and anti-inflammatory protein, which is associated with the development of both health problems.

The physical and metabolic characteristics developed in this model, allow us an important approach to the common conditions in a case of MetS.

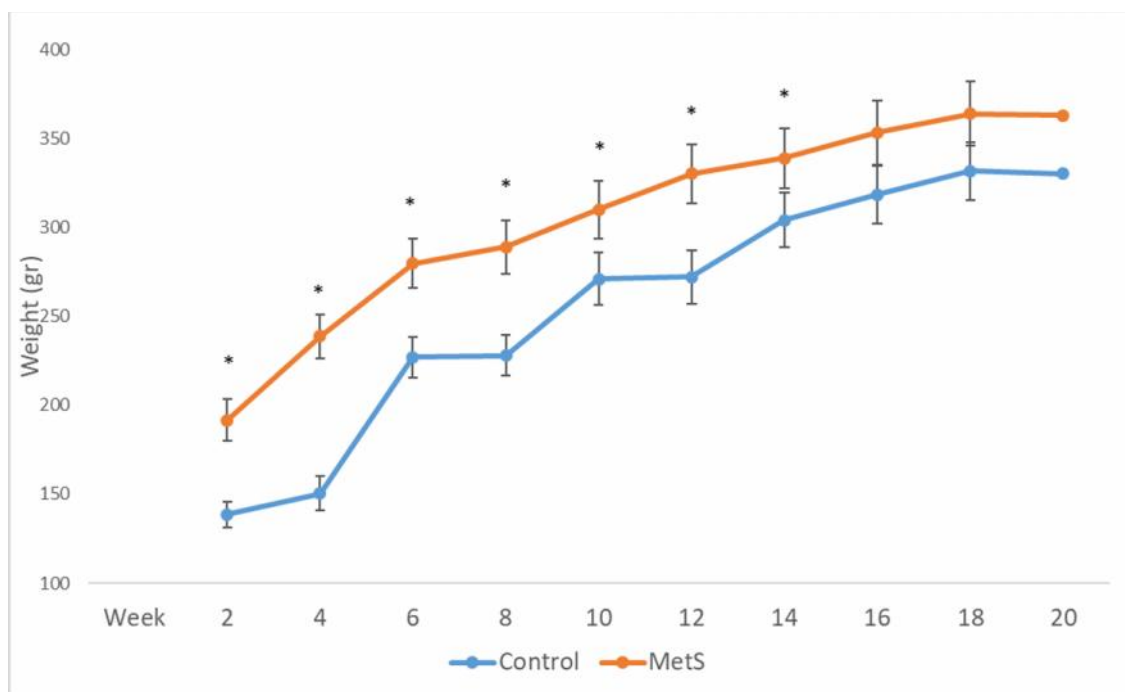


Figure 1. Body weight of rats during 20 weeks feeding. Male wistar rats were divided into the MetS group (n = 15) and the control group (n = 5). Both groups received a standard diet (2014 Teklad global). For the MetS group, a 30% sucrose solution was added to its feed and to the control group purified water *ad libitum* for 20 weeks. The weight is expressed in grams. * $P < 0.05$

Table 1. Body weight, fat total and biochemical parameters after 20 weeks feeding. Serum glucose levels, HDL and triglycerides, were measured using enzymatic methods in an automated Selectra-E equipment. Values are mean \pm SD (Control, n-5; MetS, n-15). * $P < 0.05$, ** $P < 0.01$

	Control Group	MetS Group
Body weight (g)	300 \pm 35	415 \pm 30**
Fat total (g)	20.40 \pm 1.17	37.53 \pm 1.76**
Adiposity Index	6.80 \pm 0.72	9.04 \pm 0.42*
Glucose (mg/dL)	151 \pm 11.72	158.00 \pm 6.43
Triglycerides (mg/dL)	105.00 \pm 3	205.00 \pm 10*
HDL (mg/dL)	60.61 \pm 6.66	31.74 \pm 4.00*
Total Kcal/day/100 g bw	61.05 \pm 20.06	93.03 \pm 16.91

Fat Total= Abdominal fat + Epididymal fat

Adiposity index = (Abdominal fat weight + Epididymal fat weight/Body weight) 100

Total Kcal/day/100 g bw = Kcal in drinking water + Kcal in food

References

1. The International Diabetes Federation. The IDF Consensus Worldwide Definition of the Metabolic Syndrome; International Diabetes Federation: Belgium, 2006
2. Li Y, Zhao L, Yu D, Wang Z, Ding G. Metabolic syndrome prevalence and its risk factors among adults in China: A nationally representative cross-sectional study. PLoS ONE 2018; 13 (6): e0199293.
3. Wattel F: European Committee Consensus Conference on Hyperbaric Oxygen in the Treatment of Foot Lesions in Diabetic Patients. ECHM. Glaxo-Wellcome, France, 1998.

4. Feldmeier JJ: Hyperbaric oxygen: indications and results. The Hyperbaric Oxygen Therapy Committee Report. Undersea and Hyperbaric Medical Society. Maryland, USA, 2003.
5. Bunag RD, Tomita T, Sasaki S. Chronic sucrose ingestion induces mild hypertension and tachycardia in rats. *Hypertension*. 1983;5(2):218–25.
6. Alexander Aguilera A, Hernández Díaz G, Lara Barcelata M, Angulo Guerrero O, Oliart Ros RM. Induction of Cd36 expression elicited by fish oil PUFA in spontaneously hypertensive rats. *J NutrBiochem*. 2006;17(11):760–5
7. Johnson, RJ, Segal MS, Sautin Y, Nakagawa T, Feig DI, Kang DH, Gersch MS, Benner S, Sanchez-Lozada LG. Potential role of sugar (fructose) in the epidemic of hypertension, obesity and the metabolic syndrome, diabetes, kidney disease, and cardiovascular disease. *Am J Clin Nutr* 2007; 86:899-906.
8. Oliart RR, Torres-Marquez ME, Badillo A, Angulo GO. Dietary fatty acids effects on sucrose-induced cardiovascular syndrome in rats. *J NutrBiochem* 2001; 4:207–12.
9. Teff KL, Grudziak J, Townsend RR, Dunn TN, Grant RW, Adams SH, et al., Endocrine and metabolic effects of consuming fructose- and glucose-sweetened beverages with meals in obese men and women: influence of insulin resistance on plasma triglyceride responses, *J Clin Endocrinol Metab* 2009; 94(5):1562–9.
10. Rezvani R, Cianflone K, McGahan JP, Berglund L, Bremer AA, N.L. Keim, et al., Effects of sugar-sweetened beverages on plasma acylation stimulating protein, leptin and adiponectin: relationships with metabolic outcomes. *Obesity* 2013; 21(12):2471–80.
11. Meglio DP, Mattes RD. Liquid versus solid carbohydrate: effects on food intake and body weight. *International journal of obesity and related metabolic disorders, J Int Assoc Study of Obes* 2000; 24:794–800.
12. Cornier MA, Dabelea D, Hernandez TL, Lindstrom RC, Steig AJ, Stob NR, Van Pelt RE, Wang H, Eckel RH. The metabolic syndrome. *Endocr Rev* 2008; 29(7):777–822
13. Ide T, Kobayashi H, Ashakumary L, Rouyer IA, Takahashi Y, Aoyama T, Hashimoto T, Mizugaki M. Comparative effects of perilla and fish oils on the activity and gene expression of fatty acid oxidation enzymes in rat liver. *BiochimBiophys Acta* 2000;1485(1):23–35
14. de Santa M, Olalla L, Sánchez-Muñoz FJ, Vaquero MP. N-3 fatty acids in glucose metabolism and insulin sensitivity. *Nutr Hosp* 2009; 24(2):113–127
15. Lin DS, Connor WE. Are the n-3 fatty acids from dietary fish oil deposited in the triglyceride stores of dipose tissue? *Am J Clin Nutr* 1990; 51:535–539

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