

Research Article

DOI: <http://dx.doi.org/10.22192/ijamr.2018.05.06.001>

Survey of weed flora in sugarcane fields of Tana Beles sugar development project; Ethiopia

Welday Gebreegziabher^{1*}, Ram S. Verma² and Samuel Tegene³

¹ Ethiopian Sugar Corporation, Tendaho Research and Development Center, Dubti, Ethiopia.

² Department of Plant Science, Arba Minch University, Arba Minch, Ethiopia.

³ Ethiopian Sugar Corporation, Research and Development Main Center, Wonji, Ethiopia.

*Corresponding Author: weldaygebre@gmail.com

Abstract

Survey of weed flora was carried out on 30 fields of Tana Beles Sugar Project during 2016 with the objective of evaluating the prevalence and distribution of weeds under the existing agro-ecological conditions Tana beles sugar development project. The fields were stratified into soil types (light and heavy), crop type (plant cane, ratoon and other crop (soybean)) and further in to varieties (NCO334 and N14). Three fields were surveyed in each stratum and a total of 310 samples were taken in whole survey. This was done at early growth stage (25-30 days after planting) for plant cane and soybean as well as one week before fertilization for ratoon cane fields using 0.25 m² quadrant following a pattern of inverted “W” continuously for every 2.5–3 ha. Weed species in each quadrant were collected and identified species wise. A total of 148 weed taxa were identified belonging to 31 families. The most important weed species with respect to their Important Value Index (IVI) were found to be *Brachiari aciliaris*, *Commelina benghalensis*, *Commelina diffusa*, *Commelina latifolia*, *Cynodon dactylon*, *Cyperus rotundus*, *Digitaria abyssinica*, *Eleusine indica*, *Panicum dichotomiflorum* and *Rottboellia cochinchinensis*. Moreover, all strata were infested by different weed communities leading for proper designation of weed management strategies that consider each strata independently should be made. Therefore, planning of weed control strategy in the future should take these weed diversity in to consideration.

Keywords

Important Value,
Similarity Index,
Weed flora,
Weed Species,
Weed Survey.

1. Introduction

Sugar industry development in Ethiopia has great contribution to the development of the livelihood of the society and the national economy in many ways. The demand of the society for sugar is highly increased in all directions of the country. Therefore, the government of the country had shown a high need to increase the production and productivity of sugarcane to satisfy the interest of the society in the country by initiating new projects and upgrading the existed sugar factories. But, the yield of sugarcane reduced from time to time due to different constraints. Weeds, diseases and insect pests are among the major constraints of sugarcane cultivation

in Ethiopian (Abera and Tesfaye, 2001). However, weeds are the major cause for high cost and yield reduction in sugar cane production. Although different weeds may be superficially very similar, they differ in their growth habit, reproductive habit and responses to individual control methods (Memon *et al.*, 2013). To design effective weed control measures; identification, characterization and quantification of weeds in a certain area are important steps to be followed (Firehun, 2004). Therefore, this survey was conducted with the objective of evaluating the prevalence and distribution of weed flora under agro-ecological conditions of Tana Beles Sugar Development Project plantation estate that helps to design effective weed management strategy.

2. Materials and Methods

2.1. Site Description

The study was done in Beles sugar development project, which is found in an altitude of 1119 m.a.s.l in Amhara and some part of Benshangul-gumuz regional states of Ethiopia. The average annual rainfall of Beles sub-basin is 1490 mm. The minimum and maximum temperature of the area is between 16.4 and 32.5 °C, respectively (Zelege and Netsanet, 2015). There are two main soil types i.e., heavy soil and light soil. Majority of the plantation area is covered by heavy soil.

2.2. Weed flora assessment

Purposive sampling technique was used following inverted 'W' pattern as suggested by Thomas (1985) using (0.25 m²) quadrant for every 2.5–3 ha. Number of samples per hectare was determined by species-area curve and site condition as suggested by Taye and Yohannes (1998). Plantation fields of the estate were stratified in to soil types (light and heavy) and furtherly stratified in to plant cane, ratoon cane and other crop (soybean) where prominent sugarcane varieties of the plantation (NCO334 and N14) were used for both cuttings. Three fields were surveyed in each stratum.

The survey was done during 25-30 DAP for sugarcane plant and soybean fields and a week before fertilization for ratoon fields and samples were collected using quadrant. During survey, all the weeds present in each quadrant were removed, collected and kept separately in polythene bags for species wise separation and counting. Following this, the collected weed species were then identified. For perennial grass weeds or herbaceous species, the number of shoots rather than the number of plants were counted. But, for annual grasses the tillered annual grass (rooted individuals) were counted as a single plant regardless of the number of tillers. Weeds that were difficult to identify were pressed and tagged on hard papers and then was submitted to Wonji research center for identification. The remained weeds were taken to Addis Ababa University for identification. The weeds were counted, identified and separated by species wise using the available weed identification guides (McIntyre, 1991) and were recorded in species wise

per each m² area. Nomenclature of the available weed species were also done following the flora of Ethiopia and Eritrea, volume 2, 3, 4, 6, and 7 (Hedberg *et al.*, 2003).

The data on weed survey was summarized and frequency, abundance, dominance, density, relative density, relative abundance, relative frequency, important index and similarity index values of the identified weeds were calculated using the formula described by Taye and Yohannes (1998). All the collected data were calculated by using MS-Excel.

3. Results and Discussion

3.1. Composition and diversity of weed species

A total of 145 weeds were identified at species level which were further grouped in to 31 families and 93 genera whereby each family contained 1 up to 40 weed species (Table 1 and 2). Firehun and Tamado (2007) also reported total of 180 weed taxa belonging to 40 families in wonji-shoa and Metahara sugarcane plantations.

Based on the life cycle of weeds, 47 (32%) and 101 (68%) species were found to be perennials and annuals respectively. Similar weed floral composition was reported by Yohannes *et al.* 2014 on most irrigated farms. The occurrence of annuals at higher level is due to the problem that weeding was started much delayed that most of the annual weeds were well grown. Soils of Tana Beles plantation fields contained high weed seed. Due to high moisture and continuous rainfall in the area, these weed seeds germinated but not removed on time.

Furthermore based on their morphological classification; 99 weed species (67%) were broad leaved. These were the major weeds followed by 40 species of grasses (27%) and sedges with 9 weed species (6%). Among the broad leaved species, two weeds (*Striga hermontica* (Del) Benth and *Striga aspera* (Wild) Benth) were parasitic weeds. In Ethiopia, *Striga hermontica* was reported to be prevalent in the sugarcane plantations of Fincha (Birhanu, 1993).

The survey also indicated that dicotyledonous species (67%) were dominant than monocotyledonous (33%) in the plantation estate. This might be due to less cultivation of the fields. Because, plantation of cane and sowing of soybean in the study area were done after one time cultivation followed by furrowing. Hyvonen *et al.* (2003) also reported that low input cultivations expected to favor the species numbers and abundance of dicotyledonous weeds.

Out of 31 families 14 were monotypic, viz., contained only one species. These are Acanthaceae, Boraginaceae, Caesalpiniaceae, Capparidaceae, Labiatae, Malvaceae, Plantaginaceae, Portulacaceae, Primilaceae, Robiaceae, Sphenocleaceae, Spindaceae,

Verbenaceae and Zygophyllaceae. Nigaraju *et al.* (2014) also reported Among these, Boraginaceae, Caesalpiniaceae, Robiaceae, Spindaceae, Verbenaceaeana, Zygophyllaceae as monotypic weed families in India.

Based on the number of taxa contained, 11 dominant families were identified, viz., Gramineae (40), Compositae (22), Fabaceae (11), Convolvulaceae (9), Cyperaccea (9), Amaranthaceae (7), Euphorbiaceae (7), Solanaceae (6), Commelinaceae (4), Ayzooaceae (3) and Cucurbitaceae (3) constituted a total of 121 weed species that accounts 81.75% of the total weed flora (Table 2).

Table 1: Order, Family, Richness and relative diversity of weeds in sugarcane and soybean fields

Order	Family	Richness	Relative diversity
1	Acanthaceae	1	0.68
2	Amaranthaceae	7	4.73
3	Ayzoaceae	3	2.03
4	Boraginaceae	1	0.68
5	Caesalpiniaceae	1	0.68
6	Capparidaceae	1	0.68
7	Caryophyllaceae	2	1.35
8	Chenopodiaceae	2	1.35
9	Compositae	22	14.86
10	Commelinaceae	4	2.7
11	Convolvulaceae	9	6.08
12	Cucurbitaceae	3	2.03
13	Cyperaccea	9	6.08
14	Euphorbiaceae	7	4.73
15	Fabaceae	11	7.43
16	Poaceae	40	27.03
17	Labiatae	1	0.68
18	Liliaceae	2	1.35
19	Malvaceae	1	0.68
20	Plantaginaceae	1	0.68
21	Polygonaceae	2	1.35
22	Portulacaceae	1	0.68
23	Primilaceae	1	0.68
24	Robiaceae	1	0.68
25	Scrophulaiaceae	2	1.35
26	Sphenocleaceae	1	0.68
27	Spindacea	1	0.68
28	Solanaceae	6	4.05
29	Tiliaceae	3	2.03
30	Verbenaceae	1	0.68
31	Zygophyllaceae	1	0.68
	Total	148	100.02

Therefore, Gramineae (Poaceae) was the largest family in the plantation area with 40 species followed by Compositae and Fabaceae with 22 and 11 species respectively (Table 1 and Table 2). This might be due to the high grass weed seed bank of soil in the plantation fields and they are in the same family with

sugarcane. This is similar with the result of Firehun *et al.* (2008) as they reported Poaceae (Graminae), Fabaceae, Asteraceae (Compositae), Euphorbiaceae and Convolvaceae as the dominant families that accounted for 51% of the total weed families in Fincha sugarcane plantation of Ethiopia

Table 2: Weed family, their richness and proportion percentage of eleven diverse families during the study period

Order	Family	Richness	% of total flora
1	Amaranthaceae	7	4.73
2	Ayzoaceae	3	2.03
3	Compositae	22	14.86
4	Commelinaceae	4	2.7
5	Convolvulaceae	9	6.08
6	Cucurbitaceae	3	2.03
7	Cyperaceae	9	6.08
8	Euphorbiaceae	7	4.73
9	Fabaceae	11	7.43
10	Gramineae/ Poaceae	40	27.03
11	Solanaceae	6	4.05
	Total	121	81.75

It was also reported that weeds belonging to the families Poaceae, Asteraceae and Fabaceae were the most important in arable fields of eastern Ethiopia (Tamado and Milberg, 2000). Among the dominant weed families in TanaBeles sugarcane plantation, four of them (Poaceae, Asteraceae, Euphorbiaceae, and Fabaceae) were also recorded as the most important weed families in sugarcane plantations of Coimbatore (India), Louisiana, Mauritius, and China (Lianming and Chuxiong, 2003). These families are very rich in species diversity so it is usual that they contain many weeds.

Hidalgo *et al.* (1990) reported that flora diversity is estimated to be high if the average number of species per field were greater than 19. But, in this study, diversity of flora in each field varied from 21 to 57 (mean= 39) which is high. This might be due to high soil weed seed bank, high and frequent rainfall with warm weather conditions and high fertilization. Moreover, Pulschen (1990) described the botanical family to be regarded as highly diversified; it should contain more than five species. In this study, 8 families were most diversified and contributed about 111 species which is 74.99%. The families Compositae, Convolvulaceae, Cyperaceae, Fabaceae and Poaceae were the richest taxa accounted for 91 species which is above half (61.48%) of the whole flora in the plantation site (Table 1 and Table 2).

At the genus-level, diversity was almost three times higher than that of at the family level. From the recorded 148 taxa of 93 genera, 10.76% of them contained three to seven species; the genus *Cyperus* being the most diverse taxa with seven species followed by *Brachiaria* and *Amaranthus* with each six species and *Euphorbia* with five species (Appendix 1). In Ethiopia the prevalence of *Cyperus* species was first reported by Holm *et al.* (1977). These species were found as the most dominant species in almost all the surveyed soil types of sugarcane fields in Ethiopia (Firehun, 2004).

3.2. Frequency, abundance and dominance of weeds

According to their IVI in descending order, the top 10 weeds were: *Eleusine indica*, *Cyperus rotundus*, *Commelina benghalensis*, *Commelinalatifolia*, *Panicumdichotomiflorum*, *Commelina diffusa*, *Brachiariaciliaris*, *Digitaria abyssinica*, *Rottboellia cochinchinensis* and *Cynodon dactylon* in the plantation site irrespective of soil type, crop type, sugarcane crop types and cane varieties (Table 3). Among these weeds, *Cyperus* species and *Commelina latifolia* were reported as major weed species having a higher dominance level in Wonji-Shoa sugarcane plantation estate irrespective of soil and crop types (Firehun and Tamado, 2007).

Table 3: Frequency, Abundance, Dominance, Density, Relative Frequency, Relative Abundance, Relative Density and Important value index of most important weeds at sugarcane and soybean fields

Species	TS	F	A	D	De	RF	RDe	RA	IVI
<i>Acalyphacrenata</i> A.Rich	310	3.87	0.34	1.29	0.34	1.09	1.29	1.29	3.67
<i>Achyranthesaspera</i> (L)	310	5.48	0.58	2.2	0.58	1.55	2.19	2.19	5.94
<i>Allium vineale</i>	310	6.13	0.22	0.82	0.22	1.73	0.82	0.82	3.37
<i>Amaranthusretroflexes</i> (L).	310	4.84	0.29	1.11	0.29	1.37	1.1	1.1	3.57
<i>Bidensbiternata</i> (L).	310	3.55	0.21	0.81	0.21	1	0.81	0.81	2.62
<i>Brachiariaciliaris</i>	310	6.77	1.35	5.15	1.35	1.91	5.14	5.14	12.18
<i>Brachiariacrugalli</i>	310	3.55	0.65	2.49	0.65	1	2.48	2.48	5.95
<i>Brachiarapaspaloid</i>	310	4.19	0.71	2.71	0.71	1.19	2.7	2.7	6.58
<i>Centrosemapubescens</i>	310	4.19	0.39	1.48	0.39	1.19	1.47	1.47	4.13
<i>Commelinabenghalensis</i> (L).	310	20.65	1.61	6.15	1.61	5.83	6.13	6.13	18.09
<i>Commelinadiffusa</i>	310	8.39	1.3	4.96	1.3	2.37	4.94	4.94	12.25
<i>Commelinalatifolia</i> A.Rich.	310	9.03	1.45	5.55	1.45	2.55	5.53	5.53	13.61
<i>Cynodondactylon</i> (L.) Pers.	310	8.39	0.85	3.24	0.85	2.37	3.22	3.22	8.82
<i>Cyperusesculentus</i> (L)	310	11.29	0.1	0.38	0.1	3.19	0.38	0.38	3.95
<i>Cyperusrotundus</i> (L).	310	30.32	2.8	10.69	2.8	8.57	10.65	10.65	29.87
<i>Digitariaabyssinica</i> (A. Rich) Stapf.	310	7.74	1.22	4.66	1.22	2.19	4.65	4.65	11.48
<i>Digitariasanguinalis</i> (L.) Scop.	310	2.9	0.39	1.49	0.39	0.82	1.48	1.48	3.79
<i>Echinochloacolona</i> (L.) Link.	310	2.26	0.3	1.16	0.3	0.64	1.15	1.15	2.94
<i>Eleusineindica</i> (L.) Gaertn.	310	26.45	3.15	12.04	3.15	7.47	11.99	11.99	31.45
<i>Fallopiaconvolverus</i>	310	5.48	0.61	2.33	0.61	1.55	2.32	2.32	6.18
<i>Guizotiascabra</i> (Vis) Chiov.	310	12.9	0.46	1.76	0.46	3.65	1.75	1.75	7.15
<i>Hygrophillaauriculata</i> (Schm) Heine.	310	10.97	0.6	2.29	0.6	3.1	2.28	2.28	7.66
<i>Lantana camara</i> (L.)	310	6.13	0.38	1.44	0.38	1.73	1.43	1.43	4.6
<i>Leucasmartincensis</i> (Jacq.) Ait. F.	310	5.81	0.18	0.7	0.18	1.64	0.7	0.7	3.04
<i>Nicandraphysalodes</i> (L.) Gaertn.	310	8.71	0.26	1	0.26	2.46	0.99	0.99	4.45
<i>Panicumdichotomiflorum</i> (L).	310	7.1	1.4	5.34	1.4	2.01	5.32	5.32	12.64
<i>Phalarisparadoxa</i> (L).	310	5.48	0.22	0.85	0.22	1.55	0.85	0.85	3.24
<i>Puerariaphaseoloids</i>	310	6.45	0.08	0.31	0.08	1.82	0.31	0.31	2.44
<i>Rottboelliacochinchinensis</i> (Lour.) W. D.	310	11.29	0.84	3.19	0.84	3.19	3.17	3.17	9.54
<i>Stepharniaabysinica</i> (Qu. Dill A. Roch) Walper	310	8.06	0.2	0.75	0.2	2.28	0.75	0.75	3.77
<i>Strigahermontica</i> (Del)Benth	310	4.84	0.26	1	0.26	1.37	0.99	0.99	3.35
<i>Vicia sativa</i> (L).	310	7.1	0.11	0.42	0.11	2.01	0.42	0.42	2.84
<i>Xanthium strumarium</i> (Mill.) Torrey	310	3.87	0.25	0.97	0.25	1.09	0.97	0.97	3.03

Where, TS =Total Sample, F = frequency, A = Abundance, D= Dominance, De = Density, RF= Relative Frequency, RDe= Relative Density, RA= Relative Abundance
IVI = Important value index

Moreover, the most frequent weed species recorded which accounted >8% frequency values were: *Commelina benghalensis*, *Commelina diffusa*, *Commelina latifolia*, *Cynodon dactylon*, *Cyperus esculentus*, *Cyperus rotundus*, *Eleusine indica*, *Guizotia scabra*, *Hygrophilla auriculata*, *Nicandra physalodes*, *Rottboellia cochinchinensis* and *Stephania abyssinica*. The abundant weed species having greater than 1.2 abundance values were *Brachiaria ciliaris*, *Commelina benghalensis*, *Commelina diffusa*, *Commelina latifolia*, *Cyperus rotundus*, *Digitaria abyssinica*, *Eleusine indica* and *Panicum dichotomiflorum* (Table 3).

Furthermore, the top ten dominant weeds in the plantation estate were: *Brachiaria ciliaris*, *Commelina benghalensis*, *Commelina diffusa*, *Commelina latifolia*, *Cynodon dactylon*, *Cyperus rotundus*, *Digitaria abyssinica*, *Eleusine indica*, *Panicum dichotomiflorum* and *Rottboellia cochinchinensis* which are also the top ten crowdedly found weed species (Table 3). Among these weeds, *Cyperus* species and *Commelina latifolia* were major weed species having a higher dominance level in Wonji-Shoa sugarcane plantation estate irrespective of soil and crop types (Firehun and Tamado, 2007). They also reported *Cyperus* species and *Rottboellia cochinchinensis* as the dominant weed species in Metahara sugarcane plantation. Differences were observed in abundance, frequency and dominance

values in this study due to their difference in soil, irrigation technology used, availability of weed seed bank in soil and fertilization, practices.

3.3. Similarity index value of weeds in the plantation estate

According to the description given by Taye and Yohannes (1998), if different crop types/soil types/locations have similar weed community (species composition); they would require similar weed management option. But, if crop or soil types differ in species composition, different kinds of weed control methods are required. Unger (1991) also stated that if the similarity index value is greater than 60%, it is assumed that the two locations, crops types, cane crop types and varieties are similar in species composition and hence the same weed control method can be adopted. However, if the similarity index value is below 60%, it is assumed that the two crops or soil types have different weed communities.

Accordingly, similarity indices of weed community of the study area in different soil types of different crop types and different cane varieties in the fields of the plantation ranged between 6.12 % and 48.39 % (Table 4). Thus, the similarity index values of the weed communities of all strata in both light and heavy soil types lay in much lower than 60%.

Table 4: Similarity index (%) of weed community in different soil types of TanaBeles sugarcane plantation on different crop types and cane varieties

Soil type/crop type and variety	Crop type					
	Sugarcane crop				Soybean crop	
	Plant cane		Ratoon cane			
	NCO334	N14	NCO334	N14		
Light Soil						
Plant cane	NCO 334	100	30.43	10.91	18.37	27.12
	N14	30.43	100	27.42	28.33	32.35
Ratoon cane	NCO 334	10.91	27.42	100	41.18	23.53
	N14	18.37	28.33	41.18	100	22.92
Soybean		27.12	32.35	23.53	22.92	100
Heavy Soil						
Plant cane	NCO 334	100	48.39	23.68	6.12	10.71
	N14	48.39	100	17.19	19.4	19.74
Ratoon cane	NCO 334	23.68	17.19	100	25	21.62
	N14	6.12	19.4	25	100	28.21
Soybean		10.71	19.74	21.62	28.21	100

In the other case, the similarity index (%) of weed community of different crop types and varieties of different soil types of the plantation fields ranged

between 14.29% and 44.68% (Table 5) which is also below 60%.

Table 5: Similarity index (%) of weed community in different crop types and varieties of Tana Beles sugarcane plantation in different soil types

Crop type and varieties/Soil type	Soil type	
	Light soil	Heavy soil
Plant cane		
NCo334		
Light soil	100	21.05
Heavy soil	21.05	100
N14		
Light soil	100	47.37
Heavy soil	47.37	100
Ratoon cane		
Nco334		
Light soil	100	14.29
Heavy soil	14.29	100
N14		
Light soil	100	25.71
Heavy soil	25.71	100
Soybean		
Light soil	100	44.68
Heavy soil	44.68	100

Generally, the similarity index values of all strata of the study area fall between 6.12% and 48.39%. Hence, the result showed that all the two crop types (sugarcane and soybean), soil types (light and heavy soils), the two sugarcane crop types (plant cane and ratoon cane) and the two sugarcane varieties (NCO334 and N14) have different weed communities viz. SIV < 60%. This leads for adoption of different weed management methods in the plantation site. Weed composition could change depending on some factors during a long period. Planting time and techniques, soil management, harvest time, fertilization, chemical and mechanical weed control methods are the main factors that influence weed incidence (Albrecht, 1995). In this study, the difference of weed communities within the same soil types of the same cane crop types of different cane varieties mainly attributed to the ability of weed hindrance capacity of the N14 as compared to NCO334 (Firehun *et al.*, 2013) and the soil weed seed bank.

4. Conclusion

Weeds are constant components of agro-ecosystem that have harmful effect on crop quality and quantity. So, it is a must to have accurate information on the species of weeds, their frequency, uniformity, density, coverage, growth habit and phenology. The study has ranked the most abundant and aggressive weed species in the plantation fields of Tana Beles sugar development project. A total of 148 weed taxa belonging to 31 families were recorded. Among them, 145 weeds were identified at species level while three seedlings were identified at generic level. The plantation is dominated by broadleaf weeds followed by grasses and sedges. Frequency, abundance, dominance and similarity indices constitute substantial criteria of an exact assay of weed infestation. Accordingly, due care should given to properly manage these important weeds.

Moreover, the result shown that all the strata have different weed communities. Therefore, proper designation of weed management strategies that consider each strata independently should be made. It is also recommended that survey and identification of weed flora needs to be done regularly at a certain time interval to identify the flora shift and newly introduced weeds in the plantation estate.

Acknowledgments

The authors are grateful to the financial grant of the Ethiopian Sugar Corporation, Research and Development Main Center. Many thanks are similarly forwarded to Arba Minch university and Oyda Woreda administration office for their technical advice and financial support respectively. Warmly gratitude also goes for Mr. Hadush Hagos (ESC, RDD), Kidane Tesfamicael (BRDCM), Jemal Tola (Pawi ARC, plant protection researcher) and Mrs. Ager Demise (Pawi ARC, laboratory technician) for their heartfelt collaboration and support in all aspects during the study. The staff of Beles RDC are also duly acknowledged for their supports.

References

- Abera, T. and Tesfaye H. (2001). Review of sugarcane research in Ethiopia II. Crop protection (1970-1998). Ethiopian sugar industry support centre. Research and training service, Wonji.
- Albrecht, H. (1995). Changes in Arable weed flora of Germany during the last five decades. 9th EWRS Symposium, Buda pest. 1:41-48.
- Birhanu Abreha. (1993). *A survey for identification of weed flora in Fincha sugar project site* (unpublished).
- Firehun Y., Yohannes Z. & Leul M. (2008). Study on weed composition and flora shift in Fincha sugarcane plantation. *Ethiopian Journal of Weed management*, 2: 31-43.
- Firehun, Y. & Tamado, T. (2007). Qualitative and quantitative assessments of weeds on sugarcane plantations of Wonji-Shoa and Metahara. *Ethiopia Journal of Weed Management*. 1(1), 1-14.
- Firehun, Y. (2004). *Weed flora in Wonji-Shoa and Metahara sugarcane cane plantations as influenced by some environmental and crop management practices*. M.Sc. Thesis. Alemaya University.
- Firehun, Y., Tamado, T., Abera, T. & Yohannes, Z. (2013). Weed Interference in the Sugarcane (*Saccharum officinarum* L.), Plantations of Ethiopia. *Agriculture, Forestry and Fisheries*. 2 (6): 239-247.
- Hedberg, I., Edwards, S. & Sileshi, Nemomissa. (2003). Flora of Ethiopia and Eritrea (Apiaceae to Rubiaceae), Vol. 4. The National Herbarium, Addis Ababa, Ethiopia and Department of systematic Botany, Uppsala, Sweden.
- Hidalgo, B., Saavedra, M. & Garica L. Torres. (1990). Weed flora of dryland crops in Coldeba region (Spain). *Weed Research*, 30: 309-318.
- Holm, L. G., Plucknett, D. L., Pancho, J. V. & Herberger, J. P. (1977). *The World's Worst Weeds-Distribution and Biology*. The University Press of Hawaii, pp. 609.
- Hyvonen, T., Ketoja, E., Salonen, J., Jalli, H. & Tianen, J. (2003). Weed species diversity and community composition in organic and conventional cropping of spring cereals. *Agriculture, Ecosystems & environment*, 97: 131-149.
- Lianming, C. & Chuxiong, G. (2003). *Sugarcane pests and their control*. Guangzhou Sugarcane Industry Research Institute. Guangzhou, China. p. 62.
- McIntyre, G.E. (1991). *Weeds of sugarcane in Mauritius: Their description and control*. King Keong Printing, Singapore.
- Memon, R. A., Roza Bhati, G., Khalid, S. H., Mallah, A. & Ahmed, S. H. (2013). Illustrated weed flora of wheat crop of Khairpur district, Sindh. *Pakistan Journal of botany*, 45(1):39-47.
- Nigaraju, N., Bandar, U., Rao, V. & Tarakeswara Naidu, M. (2014). Phytosociological Studies on Weed Species of Sugarcane Fields in Visakhapatnam District, Andhra Pradesh, India. *Indian International Journal Advanced Research Science Technology*, 3: 23-28.
- Pulschen, L. (1990). Compositional, synecology and sociological structure of the agrestal flora in Shewa province. *Ethiopia Angew. Botanik*, 64: 445 - 456.
- Tamado, T. & Milberg, P. (2000). Weed flora in arable fields of eastern Ethiopia with emphasis on the occurrence of *Parthenium hysterophorus* L. *Weed Research*. 40: 507 - 521.
- Taye, T. & Yohannes. (1998). Qualitative and quantitative determination of weeds in teff in west Shewazone. EWSS, Addis Ababa. In: Fassil, Reda. & Tanner, D. G. (Eds.). *Arem* 4: 46 - 60.
- Thomas, A.G. (1985). Weed survey system used in Saskatchewan for cereals and oilseed crops. *Weed Science*. 33:34-43.
- Unger, J. (1991). *Weed and weed control*. PLSC 541. A lecture note. Alemaya University of Agriculture, Alemaya, Ethiopia.

Yohanna M., Kwaga, John, M., William, B, Ndahi, Ibrahim, B., John, M. Peter & Hassan Sule. (2014). Survey of weeds on irrigated farms in Mubi area, Semi Arid Ecology, Nigeria. *International Journal of Innovative and Applied research*, 2:7-11.

Zelege, T. & Netsanet, A. (2015). Soil fertility assessment for fertilizer recommendation for sugarcane plantations at TanaBeles Sugar Development Project in Ethiopia. *African Journal of Agricultural Science and Technology*. 3(11):244-248.

Appendices

Appendix 1: Weed species found in fields of TanaBeles sugarcane plantation

Family	Species	LC	GH	Weed category
Acanthaceae				
	<i>Hygrophilla auriculata</i> (Schm) Heine.	A	Hs	Broad leaf
Amaranthaceae				
	<i>Amaranthus caudatus</i> (L.)	A	He	Broad leaf
	<i>Amaranthus graecizans</i> (L.)	A	He	Broad leaf
	<i>Amaranthus hybridus</i> (L.)	A	He	Broad leaf
	<i>Amaranthus palmeri</i>	A	He	Broad leaf
	<i>Amaranthus retroflexus</i> (L.)	A	He	Broad leaf
	<i>Amaranthus spinosus</i> (L.)	A	He	Broad leaf
	<i>Celosia trigyna</i> (L.)	A	He	Broad leaf
Ayzoaceae				
	<i>Achyranthes aspera</i> (L.)	A	He	Broad leaf
	<i>Trianthema portulacastrum</i> (L.)	A	Hp	Broad leaf
	<i>Trianthema triquetra</i> Willd.	A	He	Broad leaf
Boraginaceae				
	<i>Trichodesma zeylanicum</i> (L.) R. Br.	A	He	Broad leaf
Caesalpinaceae				
	<i>Caesalpinia decapetala</i> (Roth) Alston	A	He	Broad leaf
Capparidaceae				
	<i>Cleommono phylla</i> (L.)	A	He	Broad leaf
Caryophyllaceae				
	<i>Corrigiola litoralis</i> (L.)	P	Hp	Broad leaf
	<i>Spergula arvensis</i> (L.)	P	Hp	Broad leaf
Chenopodiaceae				
	<i>Centrosoma pubsense</i> (L.)	A	He	Broad leaf
	<i>Chenopodium album</i> (L.)	A	He	Broad leaf
Compositae				
	<i>Acanthospermum hispidum</i> DC.	A	Hp	Broad leaf
	<i>Ageratum conyzoides</i> (L.)	A	He	Broad leaf
	<i>Bidens biternata</i> (L.)	A	He	Broad leaf
	<i>Bidens pilosa</i> (L.)	A	He	Broad leaf
	<i>Conyza bonariensis</i> (L.) Cronq.	A	He	Broad leaf
	<i>Cotula abyssinica</i> Sch. Bip. ex. A. Rich	A	He	Broad leaf
	<i>Crassocephalum rubens</i> (Jacq.) S. Moore.	A	He	Broad leaf
	<i>Flaveria trinervia</i> (Spreng.) C. Mohr.	A	He	Broad leaf
	<i>Galinsoga parviflora</i> Cav.	A	He	Broad leaf

	<i>Guizotia abyssinica</i>	A	He	Broad leaf
	<i>Guizotia scabra</i> (Vis) Chiov.	A	He	Broad leaf
	<i>Launae acornuta</i> (Oliv. et Hiern) C. Jeffery.	P	Hr	Broad leaf
	<i>Mikimia macrontha</i>	P	Hv	Broad leaf
	<i>Selamum indicum</i> (L.)	A	He	Broad leaf
	<i>Sonchus asper</i> (L.) Hill	A	He	Broad leaf
	<i>Sonchus exauriculatus</i> (Olive. &Hiem.)O.Hoffm.	P	He	Broad leaf
	<i>Sonchus oleraceus</i> (L.)	P	He	Broad leaf
	<i>Spilanthes macraei</i> Hook. etArn.	A	He	Broad leaf
	<i>Spilanthes mauritiana</i> (Rich. Ex pers)DC	A	He	Broad leaf
	<i>Tagetes minuta</i> (L.)	A	He	Broad leaf
	<i>Tussilago farafara</i>	A	He	Broad leaf
	<i>Xanthium strumarium</i> (Mill.) Torrey	A	He	Broad leaf
Commelinaceae				
	<i>Commelina benghalensis</i> (L.)	A	Hs	Broad leaf
	<i>Commelina diffusa</i> BurmF.	A	Hs	Broad leaf
	<i>Commelina latifolia</i> A. Rich.	A	Hs	Broad leaf
	<i>Commelina subulata</i> Rott.	A	Hs	Broad leaf
Convolvulaceae				
	<i>Convolvulus arvensis</i> (L.)	P	Hc	Broad leaf
	<i>Convolvus spp.</i>	P	Hc	Broad leaf
	<i>Fallopiaconvolverus</i>	P	Hc	Broad leaf
	<i>Ipomoea qcquatica</i> Forssk	P		Broad leaf
	<i>Ipomoea cordofana</i> (Desr.) Choisy	P	Hp	Broad leaf
	<i>Ipomoea eriocarpa</i> R. Br.	P	Hp	Broad leaf
	<i>Ipomoea sinensis</i> Choisy	P	Hp	Broad leaf
	<i>Stepharnia abyssinica</i> (Qu. Dill A. Roch) Walper	P	Hc	Broad leaf
	<i>Zeneria Scabra</i> (L.Fil) Sonder	P	Hc	Broad leaf
Cucurbitaceae				
	<i>Cucurbit apepo</i> (L.)	A	Hc	Broad leaf
	<i>Langenaria siceraria</i>	A	Hc	Broad leaf
	<i>Momordica charantia</i>	A	Hc	Broad leaf
Cyperaceae				
	<i>Cyperus assimilis</i> Steud.	A	T	Sedge
	<i>Cyperus brevifolius</i>	P	R	Sedge
	<i>Cyperus compressus</i>	P	R	Sedge
	<i>Cyperus esculentus</i> (L.)	P	R	Sedge
	<i>Cyperus rigidifolius</i> Steud.	P	R	Sedge
	<i>Cyperus rotundus</i> (L.)	P	R	Sedge
	<i>Cyperus siberianus</i> Steudel.	P	R	Sedge
	<i>Fimbistyl islitoralis</i>	P	R	Sedge
	<i>Kyllingabulbosa</i> P. Beauv.	P	R	Sedge
Euphorbiaceae				
	<i>Acalyphacrenata</i> A. Rich	A	He	Broad leaf
	<i>Euphorbia helioscopia</i> (L.)	A	H	Broad leaf
	<i>Euphorbia heterophylla</i> (L.)	A	He	Broad leaf
	<i>Euphorbia hirta</i> (L.)	A	Hp	Broad leaf
	<i>Euphorbia indica</i> Lam.	A	He	Broad leaf
	<i>Euphorbia schimperiana</i> Scheele	A	He	Broad leaf
	<i>Ricinuscommunis</i> (L.)L.	P	S	Broad leaf

Fabaceae

<i>Acaciaspp.</i>	P	Hs	Broad leaf
<i>Cassia obtusifolia</i>	P	He	Broad leaf
<i>Centrosemapubescens</i>	P	H	Broad leaf
<i>Medicago denticulate</i>	A		Broad leaf
<i>Medicagopolymorpha (L.)</i>	A	Hp	Broad leaf
<i>Mililotus alba Medic.</i>	A	He	Broad leaf
<i>Mimosa invisa (L.)</i>	P	S	Broad leaf
<i>Mimosa pegra (L.)</i>	P	S	Broad leaf
<i>Sennaoccidentalis (L.) Link.</i>	A	Hs	Broad leaf
<i>Sesbaniaspp.</i>	A	He	Broad leaf
<i>Vicia sativa (L.)</i>	A	Hp	Broad leaf

Gramineae/ Poaceae

<i>Acroptilonrepens (L.) DC.</i>	A		Grass
<i>Brachiariaciliaeis</i>	A		Grass
<i>Brachiariacrugalli</i>	A		Grass
<i>Brachiariaeruciformis (J. E. Sm.) Griseb</i>	A	T	Grass
<i>Brachiariamutica</i>	A		Grass
<i>Brachiarapaspaloid</i>	A		Grass
<i>Brachiarareptans(L. Gard and Hubb)</i>	A		Grass
<i>Cynodondactylon (L.) Pers.</i>	P	R	Grass
<i>CynodonnlemfuensisVanderyst.</i>	P	Hp	Grass
<i>Dactylocteniumaegyptium</i>	P		Grass
<i>Digitariaabyssinica (A. Rich) Stapf.</i>	A	He	Grass
<i>Digitariaciliaris (Retz.) Koel</i>	A	T	Grass
<i>Digitariasanguinalis (L.) Scop.</i>	A	T	Grass
<i>Digitariascalarum(schweif.) Chiov.</i>	A		Grass
<i>Dinebraretroflexa (Vahl.) Panzer</i>	A	T	Grass
<i>Echinochloacolona (L.) Link.</i>	A	T	Grass
<i>Echinochloa crus-galli (L.) P. B.</i>	A	T	Grass
<i>Echinochoahaploclada(Stapf) Stapf</i>	A		Grass
<i>Eleusineindica (L.) Gaertn.</i>	A	T	Grass
<i>Eleusinemultiflora</i>	A		Grass
<i>Eragrostisaspara(Jacq.) Neels.</i>	A		Grass
<i>Eragrostiscilianensis (All.) Lut.</i>	A	T	Grass
<i>Ericholafatmensis (Hochst. et Steud.)W. D.</i>	A	T	Grass
<i>Oplismenuscompositus(L.P) Beav.</i>	P		Grass
<i>Oplismenushirtellus(L.) P.Beauv.</i>	P		Grass
<i>Panicumdichotomiflorum(L.)</i>	P		Grass
<i>Panicumrepens (L.)</i>	P	R	Grass
<i>Paspalumdistichum</i>	P		Grass
<i>Paspalumnotatum Fluegge</i>	P	T	Grass
<i>PennisetumclandestinumHochst. ExChiov.</i>	P	Hs	Grass
<i>PennisetumglabrumSteud.</i>	P		Grass
<i>Pennisetumpolystachion(L.) Schult.</i>	P		Grass
<i>Phalarisparadoxa (L.)L.</i>	A	T	Grass
<i>Poaannua(L.)</i>	A		Grass
<i>Puerariaphaseoloids</i>	P	Hc	Grass
<i>Rottboelliacochinchinensis (Lour.) W. D.</i>	A	T	Grass

	<i>Setariapumila</i> (Poir.) Roem. etSchult.	A	T	Grasss
	<i>Setariaverticillata</i> (L.) Beauv.	A	T	Grass
	<i>Sorghum arundinaceum</i> (Desv.) Stapf	A	T	Grass
	<i>Sorghum halepense</i> (L.) Pers.	A	T	Grass
Labiatae				
	<i>Leucasmartincensis</i> (Jacq.) Ait. F.	A	He	Broad leaf
Liliaceae			He	Broad leaf
	<i>Allium canadense</i> (L.)	A	He	Broad leaf
	<i>Allium vineale</i> (L.)	A	He	Broad leaf
Malvaceae				
	<i>Sidaacuta</i> Burm. F.	P	He	Broad leaf
Plantaginaceae				
	<i>Plantagolancolata</i> (L.)	P	He	Broad leaf
Polygonaceae				
	<i>Oxygonumsinuatum</i> (Meisn.) Dammer	A	Hs	Broad leaf
	<i>Polygonumnepalensis</i> Meisn.	A	He	Broad leaf
Portulacaceae				
	<i>Portulacaoleracea</i> (L.)	A	Hp	Broad leaf
Primilaceae				
	<i>Anagallisarvensis</i> (L.)	A	Hs	Broad leaf
Robiaceae				
	<i>Poederiafoetida</i> (L.)L.	P	R	Broad leaf
Scrophulaiaceae				
	<i>Strigaaspera</i> (Wild) Benth.	PAR	H	Broad leaf
	<i>Strigahermontica</i> (Del) Benth.	PAR	H	Broad leaf
Sphenocleaceae				
	<i>Sphenocleazeylanica</i> Gaerth	A	Hs	Broad leaf
Spindacea				
	<i>Cardiospermumhelicabum</i> (L.)	A	Hc	Broad leaf
Solanaceae				
	<i>Daturastramonium</i> (L.)	A	He	Broad leaf
	<i>Lycopersiconlycopersicum</i> (L.)	A	He	Broad leaf
	<i>Nicandraphysalodes</i> (L.) Gaertn.	A	He	Broad leaf
	<i>Physalis minima</i> (L.)	A	He	Broad leaf
	<i>Solanumcaroliuense</i>	P	T	Broad leaf
	<i>Solanumnigrum</i> (L.)	A	He	Broad leaf
Tiliaceae				
	<i>Corchoruschleocrusgalii</i> (L.)	A	He	Broad leaf
	<i>Corchoruspseudocapsularis</i> Schweinf.	A	He	Broad leaf
	<i>Corchorustrilocularis</i> (L.)	A	He	Broad leaf
Verbenaceae				
	<i>Lantana camara</i> (L.)	P	S	Broad leaf

Zygophyllaceae

Tribulusterrestris (L.)

A

Hp

Broad leaf

Where, A = Annuals, P= Perennials, LC = life cycle, PAR = Parasites, GH = Growth habit, H = Herb, S = Shrub, R = Rhizomatous with vegetative Propagules, T = Tufted, p= prostrate, v= vine, e=erect, c= climber, s= sprawling.

Access this Article in Online	
	Website: www.ijarm.com
	Subject: Agricultural Sciences
Quick Response Code	
DOI:10.22192/ijamr.2018.05.06.001	

How to cite this article:

Welday Gebreegziabher, Ram S. Verma and Samuel Tegene. (2018). Survey of weed flora in sugarcane fields of Tana Beles sugar development project; Ethiopia. Int. J. Adv. Multidiscip. Res. 5(6): 1-13.

DOI: <http://dx.doi.org/10.22192/ijamr.2018.05.06.001>